

1 **Photo-Fenton Degradation Kinetics of Low Ciprofloxacin**
2 **Concentration Using Different Iron Sources and pH**

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28 Abstract

29 The aim of the present study was to compare the degradation kinetics of low (1 mg L^{-1})
30 and high (25 mg L^{-1}) concentrations of ciprofloxacin (CIP) aiming to decrease the
31 concentration of additives and evaluate the pH limitation by the use of low iron
32 concentrations and organic ligands. A parameterized kinetic model was satisfactorily
33 fitted to the experimental data in order to study the performance of photo-Fenton
34 process with specific iron sources (iron citrate, iron oxalate, iron nitrate) under different
35 pH medium (2.5, 4.5, 6.5). The process modeling allowed selecting those process
36 conditions (iron source, additives concentrations and pH medium) which maximize the
37 two performance parameters related to the global equilibrium conversion and kinetic
38 rate of the process. For the high CIP concentration, degradation was very influenced by
39 the iron source, resulting in much lower efficiency with iron nitrate. At pH 4.5, highest
40 TOC removal (0.87) was achieved in the presence of iron citrate, while similar CIP
41 conversions were obtained with oxalate and citrate (0.98 after 10 min). For the low CIP
42 concentration, much higher conversion was observed in the presence of citrate or
43 oxalate in relation to iron nitrate up to pH 4.5. This behavior denotes the importance of
44 complexation also at low dosages. Appropriate additives load ($320 \text{ } \mu\text{M H}_2\text{O}_2$; $6 \text{ } \mu\text{M Fe}$)
45 resulted in a CIP conversion of 0.96 after 10 minutes reaction with citrate up to pH 4.5.

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50 Keywords: antibiotic; emerging contaminants; citrate; oxalate; kinetic model; photo-
51 Fenton

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54 **Introduction**

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56 The pharmaceuticals represent an emerging group of organic contaminants of
57 environmental concern used in human and veterinary medicine. Their continuous use
58 associated with the inefficiency of conventional sewage treatment systems contribute
59 for the contamination of aquatic medium [1]. Many studies confirm the presence of
60 these compounds in various aquatic compartments, as surface water, sewage treatment
61 plant effluents (STP), seawater and groundwater [2-7]. Among these pharmaceuticals
62 are the antibiotics, which are therapeutic agents used in infections by microorganisms
63 which act on its growth inhibition or death. Beside its main application, the antibiotics
64 have been widely used as growth promoters in animal farming [8]. Due to the poor
65 absorption and partial metabolism by the organisms, they are excreted through the feces
66 and urine [9].

67 As a consequence, the continuous exposure of aquatic organisms to low
68 concentrations of antibiotics may lead to bacterial resistance causing the inefficiency of
69 future application of these medicines [10, 11]. Furthermore, the antibiotics may
70 decrease the efficiency of the wastewater treatment process and disrupt the microbial
71 environment in water [12].

72 Ciprofloxacin (CIP) is a fluoroquinolone broad-spectrum antibiotic, which is
73 active against gram-positive and gram-negative bacteria [13], interfering in the catalytic
74 cycle of important enzymes for the nucleic acid synthesis of bacteria [14].

75 It has been observed that the relatively high removal efficiency of ciprofloxacin
76 in sewage treatment plants (STP) is mainly due to sorption and photodegradation [15,
77 16]. Ciprofloxacin has been found in hospital effluents at concentrations varying from
78 11-99 $\mu\text{g L}^{-1}$ [17, 18], in STP influents at concentration of 0.14 $\mu\text{g L}^{-1}$ [19], effluents at

79 concentrations between 105 ng L^{-1} and $0.055 \text{ } \mu\text{g L}^{-1}$ [19, 20] and raw drinking water at
80 concentration of $0.032 \text{ } \mu\text{g L}^{-1}$ [21].

81 The advanced oxidation processes (AOP) have already ratified their efficiency
82 for the abatement of pharmaceuticals in water due to the non-selectivity of the hydroxyl
83 radical, which may be effective towards different classes of pharmaceuticals which are
84 found at low concentrations permitting its effective degradation [22-25].

85 In this concern, the photo-Fenton process is especially interesting for the
86 application to the treatment of pharmaceuticals contaminated wastewater. Its high
87 efficiency of hydroxyl radical generation by the use of iron salts and hydrogen peroxide
88 at ambient temperature and pressure under UV or solar radiation has been already
89 proved [26].

90 The photo-Fenton process is tightly pH dependent, with a maximum degradation
91 efficiency of pollutants in a narrow pH range, between 2.5 to 3.0. At values above 3.0,
92 precipitation of iron (III) hydroxides occurs reducing its interaction with H_2O_2 and
93 consequently decreasing the production of $\cdot\text{OH}$. At pH below 2.5, scavenging of $\cdot\text{OH}$ by
94 H^+ ions may also decrease the efficiency of the reaction [27]. Iron complexes have been
95 applied in order to extend the pH range used in the photo-Fenton process, since Fe(III)
96 complexes are soluble at neutral pH values. The application of organic ligands as
97 oxalate and citrate in the photo-Fenton process has been shown to increase the
98 degradation efficiency of pollutants [28-31]. Furthermore, the use of iron complexes as
99 ferrioxalate (FeOx) is especially interesting for solar applications, since it absorbs
100 strongly between 250-500 nm and has a high quantum efficiency of Fe^{2+} generation
101 [32].

102 Considering that pharmaceutical concentrations found in different environmental
103 samples are in the range of ng L^{-1} to $\text{ } \mu\text{g L}^{-1}$ it is reasonable to ponder that different

104 condition of Fenton reaction may be applied concerning pH, iron source and hydrogen
105 peroxide concentration. The hypothesis is that reducing the iron concentration,
106 precipitation is reduced permitting to apply the process at higher pH values. The
107 question is then how much may pH be increased without decreasing the process
108 efficiency. Another aspect refers to the iron species. In this concern the question is how
109 important are organic ligands when low iron concentrations are applied.

110 Considering these two questions and that photo-Fenton degradation of emerging
111 contaminants at low levels and pH values close to neutral was scarcely studied [24, 31,
112 33], the aim of the present study was to compare the degradation kinetics of high and
113 low concentrations of ciprofloxacin aiming to decrease the concentration of additives
114 and evaluate the pH limitation by the use of low iron concentrations and organic ligands
115 (citrate and oxalate).

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117 **2. Material and methods**

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119 *2.1 Reagents*

120 Monohydrated ciprofloxacin hydrochloride (98%) ($C_{17}H_{18}O_3N_3F \cdot HCl \cdot H_2O$;
121 $MM=385.8 \text{ g mol}^{-1}$) was obtained from Pharmanostra. $Fe(NO_3)_3 \cdot 9H_2O$ (Mallinkrodt)
122 was used to prepare aqueous 0.25 mol L^{-1} iron stock solution. Potassium oxalate (J.T.
123 Baker) and citric acid (Synth) were used as ligands. H_2O_2 30% (w/w) (Synth) was used.
124 Bovine liver catalase was purchased from Sigma–Aldrich. Ammonium metavanadate
125 (Vetec) 0.06 mol L^{-1} was prepared in $0.36 \text{ mol L}^{-1} H_2SO_4$ (Merck) and used for
126 hydrogen peroxide determination. Methanol (HPLC grade) and formic acid (analytical
127 grade) were purchased from J.T. Baker. Ultrapure water (Millipore Milli Q water) was

128 used for dilutions and for HPLC analysis. A 0.2 mol L⁻¹ H₂SO₄ and NaOH (Chemis)
129 solution were used for pH adjustment.

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131 2.2. Experimental degradation procedures

132 Experiments were carried out in an up flow reactor previously described [27].
133 The irradiation source was a 15 W black-light lamp with maximum emission at 365 and
134 410 nm. The irradiated volume of the reactor was 280 mL and a total volume of 500 mL
135 of CIP solution was recirculated at a flow rate of 44 mL min⁻¹ using a peristaltic pump
136 (Masterflex 7518-12). The iron complexes, iron citrate (Fecit) and ferrioxalate (FeOx)
137 were prepared *in situ* by the addition of citric acid or potassium oxalate to iron nitrate
138 solution at the molar ratio iron to ligand 1:1 and 1:3, respectively. The pH of the
139 solution was then adjusted to the desired value within the range 2.5-6.5 by addition of
140 H₂SO₄ or NaOH. After pH adjustment, appropriate volume of H₂O₂ was added to the
141 solution while magnetically stirred and it was then immediately pumped into the
142 reactor. The lamp was only switched on once the reactor was completely filled, and at
143 this point, the time started to be monitored. The initial concentration of ciprofloxacin
144 was 25 mg L⁻¹ or 1 mg L⁻¹, corresponding to 65 and 2.6 μmol L⁻¹, respectively.

145 Theoretical stoichiometric H₂O₂ dose for totally degrading 65 μmol L⁻¹ CIP
146 (high ciprofloxacin concentration) into CO₂, H₂O and inorganic acids is 3 mmol L⁻¹. In
147 these experiments hydrogen peroxide dose was fixed as 2 times the stoichiometric
148 amount, 6 mmol L⁻¹ (204 mg L⁻¹), for the three pH values and three iron sources.
149 Experiments with 8 mmol L⁻¹ hydrogen peroxide dose were also carried out at pH 2.5.
150 Over stoichiometric hydrogen peroxide concentrations are desirable in order to ensure
151 enough amount for the Fenton reaction whatever iron source is tested. For the low
152 ciprofloxacin concentration (2.6 μmol L⁻¹), the concentration of hydrogen peroxide was

153 fixed ensuring a $\text{H}_2\text{O}_2/\text{Fe}$ ratio of 50 with 2.6, 6.4, 16 $\mu\text{mol L}^{-1}$ iron (0.14, 0.36, 0.90 mg
154 L^{-1}). The lowest iron concentration was equal to the CIP concentration and then
155 increased 2.5 times. Thus the hydrogen peroxide concentrations applied were 0.13, 0.32
156 and 0.80 mmol L^{-1} (4.4, 11 and 27 mg L^{-1}), which correspond to approximately 1, 2.5
157 and 6.5 times the stoichiometric amount, respectively.

158 Table 1 summarizes the operating conditions of every experiment carried out
159 under high (H) and low (L) concentrations of ciprofloxacin and additives. Molar
160 $\text{H}_2\text{O}_2/\text{Fe}$ and $\text{H}_2\text{O}_2/\text{CIP}$ ratios are also indicated.

161

162 2.3. Chemical analysis

163 The concentration of ciprofloxacin during the experiments at high concentrations
164 was determined using reversed-phase high performance liquid chromatography (HPLC),
165 coupled to a diode array detector (DAD) (SPD-M20A). At low concentrations, a
166 fluorescence detector (FL) (RF-20A) from Shimadzu (LC 20AT Prominense) was used.
167 A C-18 column (Shim-pack CLC (M) 5 μ x 250 x 4.6 mm Shimadzu) was used in both
168 cases and the mobile phase was a mixture of methanol:formic acid 0.1% (24:76) at a
169 flow rate of 1.0 mL min^{-1} . The injection volume was 40 μL . Under these conditions,
170 retention time of ciprofloxacin was 9.6 min. The wavelength of 280 nm was used for
171 detection in HPLC-DAD system and the quantification limit was 0.325 mg L^{-1} . In the
172 HPLC-FL system, the chromatographic conditions were the same as for DAD, using the
173 wavelength of 278 nm for excitation and 445 nm for emission. The quantification limit
174 in this case was 0.0137 mg L^{-1} . The enzyme catalase was used to interrupt the Fenton
175 reaction by the decomposition of residual H_2O_2 , after pH adjustment to 6-7 for iron
176 precipitation. The samples were filtered through 0.45 μm polyvinylidene fluoride
177 (PVDF) membrane (Millipore) before HPLC analysis.

178 The mineralization of organic matter during ciprofloxacin degradation was
179 evaluated by measuring the decay of the total organic carbon concentration (TOC) using
180 a TOC analyzer (Shimadzu TOC 5000A). TOC was measured immediately after the
181 sample withdrawal and without previous treatment. The TOC content includes the
182 carbon from the target compound, from the degradation products generated during
183 irradiation and from oxalate or citrate when these ligands were used. It is important to
184 mention that the proportion of oxalate to iron was 3:1 while of citrate was 1:1, resulting
185 in the same initial carbon amount since citrate has 6 carbon atoms and oxalate 2. So the
186 initial TOC concentration when oxalate or citrate was used was 24.8 mg L^{-1} and 13.3
187 mg L^{-1} in the case of iron nitrate for high CIP concentration, which refers only to CIP
188 content.

189 When performing the degradation experiments of ciprofloxacin at 25 mg L^{-1}
190 both mineralization and the decay of ciprofloxacin concentration were measured. When
191 performing the degradation experiments of ciprofloxacin at 1 mg L^{-1} , mineralization
192 was not evaluated since the quantification limit of this technique (1 mg L^{-1}) was not
193 sufficiently low, since TOC theoretical values varied from 0.72 to 1.7 mg L^{-1} for the
194 different oxalate or citrate concentrations.

195 The residual hydrogen peroxide concentration was determined
196 spectrophotometrically (Shimadzu UV mini-1240) by measuring the absorption at 450
197 nm after reaction with ammonium metavanadate [34].

198

199 *2.4. Kinetic parameters analysis*

200 A semi-empirical model already evaluated in previous works was proposed for
201 characterizing the performance of the process under study [35]. The simplifying
202 approaches to describe the TOC evolution are:

203 ✓ 1st order kinetics, for which the rate is a function of the initial values of these
 204 factors.

205 ✓ Equilibrium state (*plateau*) is achieved under specific conditions, so, a limit
 206 concentration exists, $[TOC]^\infty$.

207 A driving force leading to the equilibrium may be proposed, which is
 208 proportional to the gap to the equilibrium, $[TOC]-[TOC]^\infty$. The degradation rate may be
 209 formulated as:

$$\frac{d[TOC]}{dt} = -k_{TOC}([TOC] - [TOC]^\infty) \quad (1)$$

210 The integration of this rate law leads to the analytical expression for the TOC
 211 evolution under given initial conditions:

$$[TOC] = [TOC]^\infty + ([TOC]^0 - [TOC]^\infty) \cdot e^{-k_{TOC} t} \quad (2)$$

212 which can be expressed in terms of maximum TOC removal (ξ , degradation
 213 attained) by the following equation:

$$\xi = \xi^{max} (1 - e^{-k_{TOC} t}) \text{ being } \xi^{max} = \frac{[TOC]^0 - [TOC]^\infty}{[TOC]^0} \quad (3)$$

214 Hence, the performance of the degradation process may be characterized by
 215 determining the two parameters of the model, ξ^{max} and k_{TOC} , which were obtained by
 216 fitting the model to the experimental data under the least squares criterion.

217 In the case of the decay of ciprofloxacin concentration, since total conversion
 218 occurs in most of the cases, the conversion after 10 min ($\xi^{10 \text{ min}}$) was used for
 219 characterizing the degradation process besides of k , first order kinetic, represented in
 220 this case as k_{CIP} .

221 3. Results and discussion

222

223 A set of experiments varying iron species, pH and concentrations of
224 ciprofloxacin and additives under blacklight radiation were carried out. Firstly the
225 degradation of ciprofloxacin (CIP) was studied at the initial concentration of 25 mg L^{-1}
226 ($65 \text{ } \mu\text{mol L}^{-1}$) using fixed concentrations of iron (0.16 mmol L^{-1} ; 9 mg L^{-1}) and
227 hydrogen peroxide (6 mmol L^{-1} ; 204 mg L^{-1}), however at different pH values (2.5, 4.5
228 and 6.5) and complexing agents (oxalate and citrate) in order to evaluate the photo-
229 Fenton degradation kinetics of the antibiotic. Experiments with $8 \text{ mmol L}^{-1} \text{ H}_2\text{O}_2$ (292
230 mg L^{-1}) and 0.16 mmol L^{-1} iron were also carried out at pH 2.5 with the aim of
231 comparing the degradation of high and low CIP concentrations with the same $\text{H}_2\text{O}_2/\text{Fe}$
232 ratio..

233 Subsequently, considering the lack of studies on photo-Fenton degradation of
234 emerging contaminants at low concentrations at pH values close to neutral and reduced
235 additives concentrations, the initial concentration of ciprofloxacin was reduced to 1 mg
236 L^{-1} ($2.6 \text{ } \mu\text{mol L}^{-1}$). Iron and hydrogen peroxide concentrations were also reduced aiming
237 to evaluate the pH limitation and to study its degradation kinetics under these low level
238 concentrations. It is important to mention that no significant change in the pH (± 0.2)
239 was observed until the end of experiment for initial pH values of 2.5 and 4.5 for both
240 high and low concentrations. Only for the initial pH 6.5 a slight increase to 7.1 was
241 observed.

242

243 *3.1. Degradation of ciprofloxacin (CIP) at high dosages*

244 When comparing the conversion of 25 mg L^{-1} ($65 \text{ } \mu\text{mol L}^{-1}$) ciprofloxacin using
245 the three iron species at pH 4.5, the benefit of using the iron complexes FeOx and Fecit
246 for the pharmaceutical conversion was clear, reaching almost total conversion after 10
247 minutes with both complexes, while in the presence of iron nitrate, only about 0.20 of

248 CIP was converted after the same time (Figure 1A). Although the conversion of
249 ciprofloxacin with either oxalate or citrate was very similar, citrate achieved higher
250 maximum TOC removal, 0.699, and with oxalate the maximum conversion achieved
251 was 0.411, while insignificant TOC removal (below 0.05) was observed when using
252 iron nitrate (Fig. 1B). The higher CIP conversion in the presence of iron citrate and
253 oxalate in relation to iron aqueous-complexes can be attributed to the higher quantum
254 yield for Fe(II) generation (Φ_{FeII}) [36, 37]. Furthermore, the quantum yield of either iron
255 aqueous-complexes as well as organic complexes depends on the solution pH.
256 According to previous works, increasing pH from 2.7 to 4.0 increases the Φ_{FeII} in
257 approximately 50% in the presence of citrate. However, in the presence of oxalate, the
258 same increase in pH causes a decrease in the Φ_{FeII} of about 50% [38].

259 It was also observed a very different H_2O_2 consumption for the three iron
260 sources as pH increases. No significant difference on H_2O_2 consumption was observed
261 at pH 2.5, while at pH 4.5, 80% were consumed after 90 min in the presence of Fecit
262 and FeOx and only 30% were consumed in the presence of iron nitrate (data not
263 shown). These results indicate the low reactivity of $\text{Fe}(\text{NO}_3)_3$ with H_2O_2 at this pH
264 condition leading to low degradation efficiency. The higher H_2O_2 consumption in the
265 presence of organic complexes is also related to the higher quantum yield of Fe(II)
266 generation when compared to iron nitrate, as previously reported [25, 39, 40].

267 The system performance can be well visualized by the rate constant of CIP
268 conversion (k_{CIP}) vs. CIP conversion after 10 min ($\xi^{10 \text{ min}}$) and compared to rate constant
269 of mineralization (k_{TOC}) vs. maximum TOC removal (ξ^{max}) chart (Fig. 2), allowing the
270 parametrical characterization of the influences of iron source and pH on the system
271 behaviour. In relation to CIP conversion after 10 min, the parameters of the model move
272 within the range $\xi^{10 \text{ min}} = [0.194, 0.991]$ and $k_{\text{CIP}} = [0.002, 0.065]$, with highest rate

273 constant observed with Fecit at pH 2.5, while very similar CIP conversions were
274 obtained with FeOx (pH 2.5 and 4.5), Fecit at pH 4.5 and nitrate at pH 2.5, however in
275 all cases with much lower rate constants than with Fecit at pH 2.5. Further increase of
276 pH to 6.5 decreased significantly both the CIP conversion after 10 min and rate constant
277 for all iron species, showing clearly the pH limitation for CIP degradation under these
278 conditions even with FeOx or Fecit complexes (Fig. 2A).

279 In relation to mineralization, the parameters of the model move within the
280 following range: $\xi^{\max} = [0.070, 0.878]$ and $k_{\text{TOC}} = [0.011, 0.050]$, with minimum values
281 corresponding to iron nitrate at pH 6.5 and maximum values to iron oxalate at pH 2.5
282 (Fig. 2B). Similar maximum TOC removal at pH 2.5 were obtained with FeOx and
283 Fecit. Iron nitrate resulted also in relatively high TOC removal at pH 2.5, however at
284 much lower rate constant. It is important to mention that in the case of iron nitrate at
285 both pH 4.5 and 6.5, the results did not fit the semi-empirical kinetic model proposed,
286 since the ξ^{\max} attained in both cases was insignificant (below 0.05).

287 Given the good results of iron citrate, the study of ciprofloxacin degradation at
288 low concentrations was focused on this complex, while for comparison some
289 experiments were also carried out with other iron sources.

290

291 3.2. Degradation of ciprofloxacin at low dosages

292 For the degradation of 1 mg L^{-1} ($2.6 \text{ } \mu\text{mol L}^{-1}$) ciprofloxacin, the iron
293 concentration was varied from 2.6 to $16 \text{ } \mu\text{mol L}^{-1}$. An experiment carried out in the
294 absence of hydrogen peroxide with $16 \text{ } \mu\text{mol L}^{-1}$ Fecit (experiment L0, Table 1) resulted
295 in a very low CIP conversion ($\xi^{10 \text{ min}}$) of 0.142 and rate constant (k_{CIP}) of 0.002,
296 indicating that the contribution of iron citrate photolysis for the degradation of the target
297 compound is not substantial. In the experiments in the presence of hydrogen peroxide,

298 although the much lower iron concentration would reduce the tendency of iron
299 precipitation during the degradation process, it is possible to observe that iron
300 complexation is still very important for an efficient degradation of CIP. When
301 comparing the CIP conversion using iron citrate with iron nitrate at $6.4 \mu\text{mol L}^{-1}$ iron
302 concentration, both at pH 2.5, much higher CIP conversion was observed with iron
303 citrate achieving 0.94 after 10 min, while with iron nitrate only 0.54 were achieved,
304 similar to the results obtained at pH 6.5 with iron citrate (Fig. 3). Despite the low iron
305 concentrations, iron nitrate promoted also low CIP conversions at pH 4.5 and 6.5,
306 achieving 0.31 and 0.20, respectively.

307 An interesting aspect is that at this iron concentration range, the efficiency of
308 CIP conversion with the use of citrate is independent of pH until pH 4.5, declining
309 considerably at pH 6.5. Similar behavior was observed with FeOx, although a slight
310 decrease of CIP conversion was observed when increasing the pH from 2.5 to 4.5 (Fig.
311 3C). This may be related to the higher Φ_{FeII} of iron citrate at pH around 4 in comparison
312 to FeOx, as previously discussed [38].

313 As already shown for high dosages (section 3.1), the system performance can be
314 well visualized in the conversion rate constant (k_{CIP}) vs. CIP conversion after 10 min
315 ($\xi^{10 \text{ min}}$), allowing the parametrical characterization of the influences of iron source and
316 pH on the system behavior (Fig. 4). The parameters of the model move within the
317 following range: $\xi^{10 \text{ min}} = [0.309, 0.989]$ and $k_{\text{CIP}} = [0.004, 0.068]$ with a satisfactory
318 model adjustment, with only one value with correlation coefficient R^2 below 0.950
319 (0.899).

320 For a low ciprofloxacin concentration ($2.6 \mu\text{mol L}^{-1}$) it is also interesting to
321 discuss the effect of the reagent amounts related to the CIP concentration, remembering
322 that in all cases $\text{H}_2\text{O}_2/\text{Fe}$ ratio was fixed at 50. A similar behavior was observed with

323 Fecit and FeOx at pH 2.5 and pH 4.5 when hydrogen peroxide concentration was 2.5
324 times the stoichiometric amount. However, using stoichiometric H_2O_2 concentration
325 ($\text{H}_2\text{O}_2/\text{CIP} = 50$), only Fecit resulted in significant CIP conversions after 10 min (0.75)
326 at pH 2.5 and 4.5, indicating that Fecit is the iron species least affected by the pH.

327 Higher $\text{H}_2\text{O}_2/\text{CIP}$ ratio ($\text{H}_2\text{O}_2/\text{CIP} = 308$) resulted in an important effect on the
328 rate constant, achieving $k_{\text{CIP}} = 0.07$, but a lower effect on CIP conversion after 10 min.
329 On the other hand, these results suggest that at intermediate concentrations, as
330 $\text{H}_2\text{O}_2/\text{CIP}$ ratio = 123, almost total conversions (higher than 0.90) are observed after 10
331 min with rate constants between 0.02 and 0.03 with Fecit up to pH 4.5. In relation to
332 oxalate, similar results are obtained only at pH 2.5.

333 The degradation of low and high CIP dosages under the same $\text{H}_2\text{O}_2/\text{CIP}$ ratio
334 (123) and $\text{H}_2\text{O}_2/\text{Fe}$ ratio (50) at pH 2.5 (experiments H10 and L4; H11 and L10; H12
335 and L13) resulted in very similar degradation behavior (data not shown), demanding
336 however, lower concentrations of iron and hydrogen peroxide at low CIP dosages.

337

338 **Conclusions**

339

340 The kinetic model applied was satisfactorily fitted to the experimental data. The
341 degradation performance of high and low concentration of ciprofloxacin was
342 characterized by TOC removal (ξ^{max} , k_{TOC}) and CIP conversion ($\xi^{10 \text{ min}}$, k_{CIP}). The
343 system performance was evaluated at different Fenton additives (concentration, iron
344 source) and pH values showing that:

- 345 • The system behaved similarly under high and low concentration.

- 346 • The pH limitation showed to be critical in the absence of organic ligands,
347 resulting in low CIP conversions with iron nitrate at both high and low dosages,
348 except at pH 2.5.
- 349 • Iron citrate permitted to achieve significant CIP conversions at pH 2.5 and 4.5,
350 being the best choice up to pH 4.5 with a H₂O₂/CIP ratio between 50 and 123.
- 351 • Iron citrate was the iron species least affected by the pH.
- 352 • Similar behavior was observed with oxalate, however only at 123 H₂O₂/CIP
353 ratio.

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355

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